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Dispersion of swimming algae in laminar and turbulent channel flows: consequences for photobioreactors

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Shear flow significantly affects the transport of swimming algae in suspension. For example, viscous and gravitational torques bias bottom-heavy cells to swim towards regions of downwelling fluid (gyrotaxis). It is necessary to understand how such biases affect algal dispersion in natural and industrial flows, especially in view of growing interest in algal photobioreactors. Motivated by this, we here study the dispersion of gyrotactic algae in laminar and turbulent channel flows using direct numerical simulation (DNS) and a previously published analytical swimming dispersion theory. Time-resolved dispersion measures are evaluated as functions of the Péclet and Reynolds numbers in upwelling and downwelling flows. For laminar flows, DNS results are compared with theory using competing descriptions of biased swimming cells in shear flow. Excellent agreement is found for predictions that employ generalized Taylor dispersion. The results highlight peculiarities of gyrotactic swimmer dispersion relative to passive tracers. In laminar downwelling flow the cell distribution drifts in excess of the mean flow, increasing in magnitude with Péclet number. The cell effective axial diffusivity increases and decreases with Péclet number (for tracers it merely increases). In turbulent flows, gyrotactic effects are weaker, but discernable and manifested as non-zero drift. These results should have a significant impact on photobioreactor design.

1. Introduction

In natural bodies of water and in industrial bioreactors, microscopic algae experience laminar and turbulent flows that play a critical role in their dispersion, proliferation and productivity [1,2]. The dispersion of passive tracers in fluid flows is well understood, particularly in the industrially relevant cases of flows in pipes and channels [3]. Taylor first realized that the dispersion of passive tracers in a pipe, caused by the combination of fluid shear and molecular diffusion, could be described in terms of an effective axial diffusivity [4]. A similar result also holds for turbulent pipe [5] and channel [6] flows. Taylor's pioneering analyses of dispersion in a pipe inspired a series of studies that placed the understanding of the dispersion of neutrally buoyant tracers on a firm footing [7,8]. Particles whose density differs from the suspending medium exhibit more complex behaviour (e.g. they can accumulate at walls in turbulent channel flows; see [9,10]).

Swimming single-celled algae are known to respond non-trivially to flows. For example, the mean swimming direction of biflagellates, such as *Chlamydomonas* and *Dunaliella* spp., is biased by imposed flows [11,12]. This bias, known as gyrotaxis, results from the combination of viscous torques on the cell body, owing to flow gradients, and gravitational torques, arising from bottom-heaviness and sedimentation [13]. In the absence of flow gradients, the gravitational torque leads cells to swim upwards on average (gravitaxis). Recent simulations and laboratory experiments have shown how inertia and

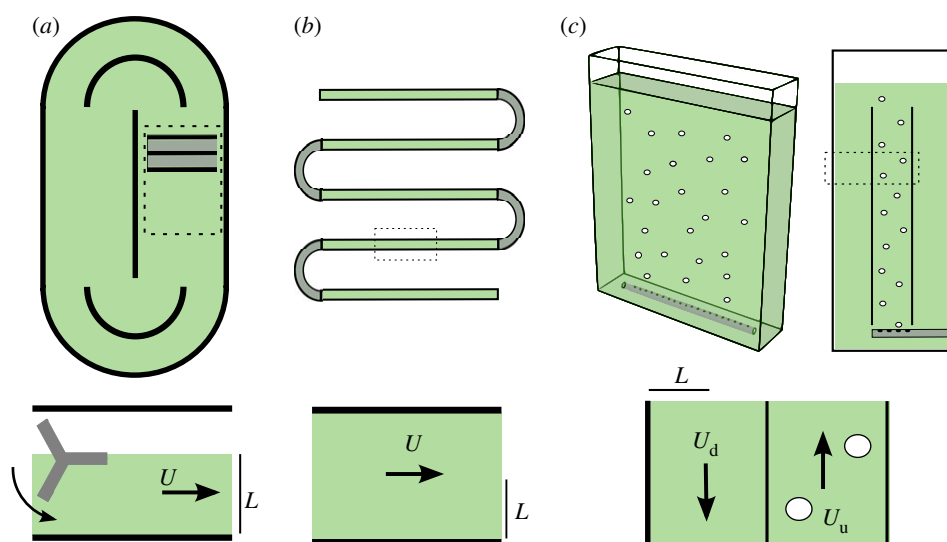


Figure 1. Examples of photobioreactors: (a) raceway pond (top view), showing the paddlewheel and guiding baffles; (b) tubular array, where suspensions are typically driven by a pump; (c) air-lift flat panel (three-dimensional view) and draft tube (cross-sectional side view), where rising bubbles generate flow as well as providing mixing and aeration. The regions highlighted with broken lines in the top diagrams are reproduced below to indicate the length scale L and magnitude U of typical flows. For the air-lift in (c), both upwelling U_u and downwelling U_d flows are shown. (Online version in colour.)

gyrotaxis can play significant roles in the formation of patchiness and/or thin layers of toxic algae in the ocean [14–16]. Gravitaxis and gyrotaxis can also couple in a complex fashion to other biases owing to chemical gradients (chemotaxis) and light (phototaxis [17]). Therefore, the fact that cells can actively swim across streamlines and accumulate in specific regions of a flow forewarns that the resultant dispersion of biased swimming algae in a complex flow field is non-trivial.

In a recent study, Bees & Croze [18] extend the classical Taylor–Aris analysis of dispersion in a laminar flow in a tube to the case of biased swimming algae. The Bees & Croze dispersion theory provides a general theoretical framework to describe the dispersion of biased micro-swimmers in confined geometries, such as pipes and channels. To make predictions for the dispersion of particular organisms, specific functional expressions for the swimming characteristics need to be obtained from microscopic models. For example, using expressions for the mean swimming orientation \mathbf{q} and diffusivity \mathbf{D} from the so-called Fokker–Planck (FP) model [12], Bees & Croze [18] found that gyrotaxis can significantly modify the axial dispersion of swimming algae (such as *Chlamydomonas* spp.) in a pipe flow relative to the case of passive tracers (or dissolved chemicals). More recently, Bearon *et al.* [19] obtained predictions for swimming dispersion in laminar flows in tubes of circular cross section by using an alternative microscopic model known as generalized Taylor dispersion (GTD), formulated for swimming algae [20,21] and bacteria [22]. The GTD model is considered superior, as it incorporates correlations in cell positions owing to cell locomotion within a local flow, as well as the gyrotactic orientational biases considered in the FP description. The qualitatively distinct predictions, however, have yet to be tested. Direct tests of the models might involve microscopic tracking of isolated cells swimming in prescribed flows. The Bees & Croze theory allows the predictions of the two models to be tested for entire populations in macroscopic experiments [23] and individual-based numerical simulations, as we shall demonstrate in this paper.

Microalgae currently play a prominent role in biotechnology. For example, they are cultured commercially as nutritious

fish and crustacean feed in aquaculture, and for high-value products that they can synthesize, such as β -carotene [24]. Microalgae also hold significant potential for future exploitation; they can provide a sustainable biofuel feedstock that does not compete for arable land with food crops. Despite significant efforts, however, current production systems remain too inefficient for algal biofuels to be commercially viable [25]. Algal culture commonly occurs in open or closed bioreactors, with production systems employing three stages: cell culture, harvest and downstream processing. Open bioreactors consist of one or more artificial ponds, quiescent or stirred, where stirring allows one to obtain greater concentrations per reactor area [24]. A common design is the raceway pond, in which rotating paddles generate flow (figure 1a). Typically, raceway ponds are shallow (depths $L = 10$ – 30 cm), which aids cell exposure to light. Typically, the flow is turbulent (speeds of 30 cm s^{-1}), facilitating light exposure and gas exchange. Pond bioreactors are relatively cheap to operate, but are limited by their low areal productivity and susceptibility to contamination. This restricts the range of viable species in open pond culture to robust, high-value extremophiles, which grow in prohibitive conditions such as high salinity or low pH. A greater variety of fast-growing algae can be cultured in closed bioreactors, where single species are propagated within sealed transparent containers. Common geometries are flat, cylindrical (columnar or tubular) and annular (figure 1b,c). As for raceways, the constraint of light penetration limits the smallest dimension to 10 cm. In tubular bioreactors, the flow speed typically is 50 cm s^{-1} , while flat panel and columnar/annular set-ups operate at 1 – 10 cm s^{-1} [26]. Although closed bioreactors allow for rapid growth conditions with small footprints, current designs are relatively expensive to manufacture, operate and maintain, and the algae are susceptible to invasion by other less-useful species.

Existing bioreactors operate under both laminar and turbulent flow conditions. The transition between the two flow regimes depends on the ratio of inertial and viscous forces in a fluid. This is expressed by the dimensionless Reynolds number $Re = UL/\nu$, where U and L are the characteristic speed and length scale of the flow, respectively, and ν is

Table 1. Non-dimensional parameters for the different cases considered in the DNS and in the analytical model predictions for the laminar case. Re is the Reynolds number of the flow, Pe and β are flow and swimming Péclet numbers, respectively, defined by equations (2.13), and α_0 is the cell eccentricity (more than zero for elongated cells, zero for spheres). L, laminar; T, turbulent.

Re (laminar/turbulent)	Pe	β	α_0
4 (L)	27	1.34, 6.7, 33.5	0
100 (L)	670	1.34, 6.7, 33.5	0 (for all β), 0.8 (for $\beta = 6.7$)
250 (L)	1675	3.35, 33.5	0
2500 (T)	16 750	33.5	0
4200 (T)	28 140	55	0, 0.8
10 000 (T)	67 000	67	0

the suspension kinematic viscosity [27]. In general, flows with $Re > 2000$ will be turbulent, though the transition to turbulence depends on geometry and particular flow conditions. Approximating the kinematic viscosity by that of water and using the flow speed and length scales above, we see that flows in air-lift reactors can be both laminar or turbulent ($Re \gtrsim 100$), while raceway ponds and tubular reactors are always turbulent ($Re > 30\,000$). The mixing properties of turbulent flows are thought to be beneficial for cell growth, although clear evidence for this is lacking. With air-lifts the mixing is caused by rising bubbles, which also provide aeration, and the flow does not need to be turbulent. Both cells and nutrients will disperse in the above flows, with cell growth and productivity depending critically on such dispersion.

Current photobioreactor designs assume that cells disperse as tracers, whether they swim or not [28]. However, as detailed above, recent studies show that the distribution and consequent dispersion of swimming cells in flows, including biotechnologically interesting species such as *Dunaliella*, should be very different from that of passive tracers [18,19]. For example, if cells disperse differently to nutrients in a reactor they will separate, which could have catastrophic consequences for growth. Inspired by the possibility of taking advantage of the peculiarities of swimmer response to flow to improve bioreactor operation, we study here the dispersion of gyrotactic swimming algae in laminar and turbulent channel flows. We carry out time-resolved direct numerical simulations (DNSs), comparing results to predictions from the Bees & Croze dispersion theory applied to channel geometry. The paper is structured as follows. In §2, we present the simulation methods and outline a derivation of the theory for the new geometry. In §3, we present simulation results for the dispersion of biased swimming algae in laminar and turbulent flows, comparing theory and simulation results for laminar flows. In §4, we interpret the results, and discuss their implications for dispersion in photobioreactors and the environment.

2. Methods

2.1. Direct numerical simulations: governing equations

We simulate the dynamics of a population of biased swimming micro-organisms, typically $N = 2 \times 10^5$ and 10^6 individuals per simulation, placed in laminar and turbulent flows. Each micro-swimmer is modelled as a spheroidal particle whose position

\mathbf{x}_i , $i = 1 \dots N$, evolves according to

$$\frac{d\mathbf{x}_i}{dt} = \mathbf{u} + V_s \mathbf{p}, \quad (2.1)$$

where V_s is the mean cell swimming speed, \mathbf{u} the local fluid velocity and \mathbf{p} the local particle orientation.

The orientation \mathbf{p} of each swimmer evolves in response to the biasing torques acting upon it. Making the realistic simplifying assumption that swimming cells have a spheroidal geometry, the reorientation rate of the organisms is defined by the inertia-free balance of gravitational and viscous torques [12,29]

$$\frac{d\mathbf{p}}{dt} = \frac{1}{2B} [\mathbf{k} - (\mathbf{k} \cdot \mathbf{p})\mathbf{p}] + \frac{1}{2} \boldsymbol{\omega} \times \mathbf{p} + \alpha_0 (\mathbf{I} - \mathbf{p}\mathbf{p}) \cdot \mathbf{E} \cdot \mathbf{p} + \Gamma_r, \quad (2.2)$$

where \mathbf{k} is a unit vector in the vertical direction; $B = \mu \alpha_\perp / (2h\rho g)$ is the gyrotactic reorientation time (h denotes the centre-of-mass offset relative to the centre-of-buoyancy, α_\perp is the dimensionless resistance coefficient for rotation about an axis perpendicular to \mathbf{p} , ρ and μ are the fluid density and viscosity, respectively); $\alpha_0 = (a^2 - b^2)/(a^2 + b^2)$ is the eccentricity of the spheroids with major axis a and minor axis b ; and, finally, \mathbf{E} is the rate of strain tensor and $\boldsymbol{\omega}$ the vorticity. The noise Γ_r is added to simulate the stochastic rotational diffusivity of a swimming cell. In the simulations it is implemented with random angular steps of magnitude $\sqrt{2d_r}$, so that cells diffuse with rotational diffusivity d_r [30,31].

The flow field \mathbf{u} is obtained by solving the Navier–Stokes equations for an incompressible viscous fluid, such that

$$\frac{D\mathbf{u}}{Dt} = -\frac{1}{\rho} \nabla p + \nu \nabla^2 \mathbf{u}, \quad \nabla \cdot \mathbf{u} = 0, \quad (2.3)$$

where $D/Dt \equiv \partial/\partial t + (\mathbf{u} \cdot \nabla)$ denotes the material derivative. Here, for simplicity, we shall assume cells to be neutrally buoyant, neglecting feedback from the particles to the flow (these effects are explored elsewhere [18]).

2.2. Direct numerical simulations: geometry, scaling and statistical measures of dispersion

We consider a standard channel geometry: two flat plates parallel to each other, infinite in extent, and separated by a gap of size $2H$. When focusing on swimming cells it would seem natural to rescale length by the plate half-width H and time by $H^2 d_r / V_s^2$, where V_s is the mean swimming speed and d_r is the cell rotational diffusivity, the characteristic time a cell swimming with V_s takes to diffuse across H with diffusivity $V_s^2 d_r^{-1}$. However, the parameter values obtained from this ‘cell-based’ rescaling are too large and thus numerically inconvenient for simulations. Thus, we adopt a ‘flow-based’ rescaling in terms of the characteristic length H and the flow-based time scale H/U_c , the time taken for a flow with centreline speed U_c

to advect a cell by H . In terms of this rescaling, the dimensionless equations of motion for a biased swimming cell are

$$\frac{d\mathbf{x}_i^*}{dt^*} = \mathbf{u}^* + v_s \mathbf{p}, \quad (2.4)$$

$$\frac{d\mathbf{p}}{dt^*} = \eta^{-1} [\mathbf{k} - (\mathbf{k} \cdot \mathbf{p})\mathbf{p}] + \frac{1}{2} \boldsymbol{\omega}^* \times \mathbf{p} + \alpha_0 (\mathbf{I} - \mathbf{p}\mathbf{p}) \cdot \mathbf{E}^* \cdot \mathbf{p} + \Gamma_r^* \quad (2.5)$$

$$\text{and } \frac{D\mathbf{u}^*}{Dt^*} = -\nabla^* p + Re^{-1} \nabla^{*2} \mathbf{u}^*, \quad \nabla \cdot \mathbf{u}^* = 0, \quad (2.6)$$

where starred quantities denote dimensionless variables. In particular, we define the dimensionless swimming speed $v_s = V_s/U_c$, the gyrotactic parameter $\eta = 2BU_c/H$ and the Reynolds number

$$Re = \frac{U_c H}{\nu}, \quad (2.7)$$

where U_c is the centreline speed, H is the channel half-width and ν the fluid's kinematic viscosity. Non-dimensionalizing the noise Γ_r also defines the dimensionless rotational diffusivity $d_r^* = d_r H/U_c$.

We adopt a Cartesian coordinate system where the mean flow in the x (streamwise) direction varies with the wall-normal coordinate y , and is independent of z (spanwise direction). We integrate equations (2.4)–(2.6) numerically (see the electronic supplementary material) to find $\mathbf{x}_i^*(t^*) = [x_i^*(t^*), y_i^*(t^*), z_i^*(t^*)]$. Enumerating the number of cells $N(\mathbf{x})$ at a given position \mathbf{x} in a bin of fixed volume ΔV , we obtain the cell concentration $c(\mathbf{x}) = N(\mathbf{x})/\Delta V$. From the cell coordinates we can further define statistical measures of the cell dispersion in a flow: the adimensional drift A_0^* with respect to the mean flow $U = (2/3)U_c$; the effective streamwise diffusivity D_e^* ; and the skewness of the cell distribution, γ . These measures of dispersion are given by

$$\left. \begin{aligned} A_0^*(t^*) &\equiv \frac{dm_1^*}{dt^*} - \frac{2}{3}, \quad D_e^*(t^*) \equiv \frac{1}{2} \frac{d}{dt^*} \text{Var}(x^*) \\ \text{and } \gamma(t^*) &\equiv \frac{m_3^* - 3m_1^*m_2^* + 2m_1^{*3}}{\text{Var}(x^*)^{3/2}}, \end{aligned} \right\} \quad (2.8)$$

where $m_p^* = (1/N) \sum_i (x_i^*)^p$ ($p = 1, 2, 3$) are the distribution moments and $\text{Var}(x^*) = m_2^* - m_1^{*2}$ is the variance of the cell distribution. The statistical measures can be transformed from the flow-based scaling with characteristic time scale H/U_c to the cell-based scaling with time scale $H^2 d_r/V_s^2$ by the transformations

$$\left. \begin{aligned} t^* &\rightarrow t \equiv \frac{t^*}{Pe}, \quad A_0^* \rightarrow A_0 \equiv A_0^* Pe \\ \text{and } D_e^* &\rightarrow D_e \equiv D_e^* Pe, \end{aligned} \right\} \quad (2.9)$$

where Pe is the cell Péclet number defined with respect to the centreline speed (see equation (2.13)). Note that the skewness γ does not depend on the scaling used. We shall present results in terms of the cell-based scaling and compare their limit at long times with analytical predictions.

2.3. Analytical theory: dispersion in laminar flows at long times

Here, we shall obtain analytical predictions for the dispersion of algae swimming in laminar channel flow that will be compared with the results from the simulations. The general Bees & Croze [18] continuum dispersion theory for biased swimmers will be applied to the new channel geometry; it is valid in the long-time limit ($t \gg H^2 d_r/V_s^2$). The derivation for channel flow is similar to the pipe flow example in Bees & Croze, so we provide an outline here. Those readers who are less mathematically inclined may skip the derivation of the results in this section.

We shall begin with the continuity equation for the cell number density n :

$$\frac{\partial n}{\partial t} = -\nabla \cdot [n(\mathbf{u} + \mathbf{V}_c) - \mathbf{D} \cdot \nabla n]. \quad (2.10)$$

The flow velocity \mathbf{u} is the solution of the Navier–Stokes equation (2.3). We adopt the same coordinate system described above for the DNS. For laminar flows such that the flow downstream is only a function of the wall-normal coordinate y , the flow velocity can be expressed as

$$\mathbf{u}(y) = u(y)\mathbf{e}_x = U[1 + \chi(y)]\mathbf{e}_x, \quad (2.11)$$

where U is the mean flow speed and $\chi(y)$ describes the variation in the streamwise direction about this mean. It is convenient to translate to a reference frame travelling with the mean flow, and non-dimensionalize length scales by H and time scales by the time to diffuse across the channel $H^2 d_r/V_s^2$. Thus, $\hat{x} = (x - Ut)/H$, $\hat{y} = y/H$, $\hat{z} = z/H$ and $\hat{t} = V_s^2 t/(H^2 d_r)$, where variables with hats denote dimensionless variables.

We assume unidirectional coupling of the cell dynamics to the flow (for bidirectional coupling owing to non-neutrally buoyant cells, see [18]): cells are biased by shear in the flow, so that the cell swimming velocity $\mathbf{V}_c = V_s \mathbf{q}$, where \mathbf{q} is the mean swimming direction, and diffusivity tensor \mathbf{D} are functions of local flow gradients. Here, we consider analytical predictions for spherical cells ($\alpha_0 = 0$), so cell orientation is only a function of vorticity, $\boldsymbol{\omega} = \nabla \times \mathbf{u} = -\chi' \mathbf{e}_z$.

Consider planar Poiseuille flow, $\chi = (1 - 3y^2)/2$ and $U = (2/3)U_c$ [32], where U_c is the centreline (maximum) flow speed. Equation (2.10) becomes

$$\frac{\partial n}{\partial t} = \nabla \cdot (\mathbf{D} \cdot \nabla n) - \frac{2}{3} Pe \chi n_x - \beta \nabla \cdot (n \mathbf{q}), \quad (2.12)$$

with

$$Pe = \frac{U_c H d_r}{V_s^2} \quad \text{and} \quad \beta = \frac{H d_r}{V_s}, \quad (2.13)$$

where hats have been dropped for clarity. Pe is a Péclet number, the dimensionless ratio of the rates of transport by the flow and swimming diffusion, and β is the ratio of channel half-width to the length a cell swims before reorienting significantly. Alternatively, we can rewrite $\beta = H V_s^2 d_r/V_s^2$ and interpret it as a 'swimming' Péclet number, the ratio of the rates of transport by swimming and diffusion. No-flow and no-flux boundary conditions will be applied to (2.12), such that

$$\mathbf{u} = \mathbf{0} \quad \text{and} \quad \mathbf{n} \cdot (\mathbf{D} \cdot \nabla n - \beta \mathbf{q} n) = 0 \quad \text{on } \Sigma, \quad (2.14)$$

where \mathbf{n} is the unit vector normal to the channel boundary Σ .

As the flow is translationally invariant along x , the mean swimming direction and diffusivity tensor are independent of x : $\mathbf{D} = \mathbf{D}(y)$, $\mathbf{q} = \mathbf{q}(y)$. This permits a treatment of dispersion using moments in a similar vein to that of Aris [7]. The p th moment with respect to the axial direction is defined as

$$c_p(y, t) = \int_{-\infty}^{+\infty} x^p n(x, y, t) dx, \quad (2.15)$$

provided $x^p n(x, y, t) \rightarrow 0$ as $x \rightarrow \pm \infty$. We denote cross-sectional averages by overbars. The cross-sectionally averaged axial moment is thus

$$m_p(t) = \bar{c}_p = \frac{1}{2} \int_{-1}^1 c_p(y, t) dy. \quad (2.16)$$

In Cartesian coordinates, equation (2.12) becomes

$$\begin{aligned} n_t &= (D^{yy} n_y - \beta q^y n + D^{xy} n_x)_y + D^{xy} n_{xy} \\ &\quad - \left(\frac{2}{3} Pe \chi + \beta q^x\right) n_x + D^{xx} n_{xx}. \end{aligned} \quad (2.17)$$

Multiplying by x^p and integrating over the length of an infinite channel, we obtain the moment evolution equation

$$c_{p,t} = (D^{yy}c_{p,y} - \beta q^y c_p - p D^{xy}c_{p-1})_y - p D^{xy}c_{p-1,y} + p(\frac{2}{3}Pe\chi + \beta q^x)c_{p-1} + p(p-1)D^{xx}c_{p-2}. \quad (2.18)$$

Averaging over the cross section and applying the no-flux boundary conditions (2.14) yields

$$m_{p,t} = p(p-1)\overline{D^{xx}c_{p-2}} - p\overline{D^{xy}c_{p-1,y}} + p\overline{[\frac{2}{3}Pe\chi + \beta q^x]c_{p-1}} \quad (2.19)$$

from which we calculate measures of cell dispersion. The drift above the mean flow, $A_0 \equiv \lim_{t \rightarrow \infty} (d/dt) m_1(t)$, is given by

$$A_0 = -\overline{D^{xy}Y_0^0} + \overline{[\frac{2}{3}Pe\chi + \beta q^x]Y_0^0}, \quad (2.20)$$

where

$$Y_0^0(y) = \exp\left(\beta \int_0^y \frac{q^y(s)}{D^{yy}(s)} ds\right) \left\{ \overline{\exp\left(\beta \int_0^y \frac{q^y(s)}{D^{yy}(s)} ds\right)} \right\}^{-1} \quad (2.21)$$

is the zeroth axial moment (normalized concentration profile). Similarly, the effective diffusivity, $D_e \equiv \lim_{t \rightarrow \infty} (1/2)(d/dt)[m_2(t) - m_1^2(t)]$, is given by

$$D_e = -\overline{D^{xy}g'} + \overline{[\frac{2}{3}Pe\chi + \beta q^x - A_0]g} + \overline{D^{xx}Y_0^0}, \quad (2.22)$$

where $g(y) = Y_0^0 \int_0^y [(D^{xy}(s)/D^{yy}(s)) - (\tilde{A}_0(s) - A_0 \tilde{m}_0(s))/D^{yy}(s)] Y_0^0(s) ds$, with $\tilde{A}_0(y) = \int_0^y [-D^{xy}Y_0^0 + [\frac{2}{3}Pe\chi + \beta q^x]Y_0^0] ds$ and $\tilde{m}_0(y) = \int_0^y Y_0^0 ds$. The weighting function $Y_0^0(y)$ controls the drift and $g(y)$ (related to the first axial moment) controls the value of the diffusivity.

To make predictions from (2.20) and (2.22), we require expressions for \mathbf{D} and \mathbf{q} from microscopic models of the statistical response of cells to flow. Two main models have been proposed for biased swimming algae: the FP model [33,34] and GTD [19–21]. For spherical cells, each model predicts how non-dimensional swimming direction \mathbf{q} and diffusivity \mathbf{D} depend on two non-dimensional quantities: $\sigma(y) = -\chi'(y)U/(2d_r H) = -\chi'(y)(1/3)(Pe/\beta^2)$, the ratio of reorientation time by rotational diffusion to that by shear (vorticity), and $\lambda = 1/(2d_r B)$, the ratio between the time of reorientation by rotational diffusion $(1/2)d_r^{-1}$ and the gyrotactic reorientation time B . The functional forms for the transport parameters $\mathbf{q}(\sigma)$ and $\mathbf{D}_m(\sigma)$, where subscripts $m = F$ and G denote the FP and GTD models, respectively, are given in appendix A [19]. In particular, the two models differ qualitatively in their predictions for the diffusivity as a function of the shear rate, and therefore they provide different predictions for cell distribution and dispersion. The dispersion predictions (2.20) and (2.22) were evaluated numerically with MATLAB (Mathworks, Natick, MA, USA) using $\mathbf{q}(\sigma)$ and $\mathbf{D}_m(\sigma)$ from the two models (see electronic supplementary material).

2.4. Parameters, simulation time and flow profiles

In simulations and theoretical evaluations we used the following cell parameters based on *Chlamydomonas augustae*: $V_s = 0.01 \text{ cm s}^{-1}$ (swimming speed), $d_r = 0.067 \text{ s}^{-1}$ (cell rotational diffusivity), and $B = 3.4 \text{ s}$ (gyrotactic reorientation time) [18]. Furthermore, we consider the flow of suspensions taken to have the same viscosity as water, $\nu = 0.01 \text{ cm}^2 \text{ s}^{-1}$. The cell eccentricity α_0 is also held fixed. The analytical theory presented above assumes that cells are spherical, $\alpha_0 = 0$, but laminar and turbulent simulations have also been performed for elongated cells with $\alpha_0 = 0.8$. With these parameters fixed, choosing the centreline speed U_c and channel width H gives the non-dimensional flow-based parameters used in the DNS: $v_s = V_s/U_c$ (dimensionless swimming speed), $\eta = 2BU_c/H$ (gyrotactic parameter), $d_r^* = d_r H/U_c$ (dimensionless rotational diffusivity) and Reynolds number $Re = U_c H/\nu$. For the test run with passive tracers an additional noise term was added to

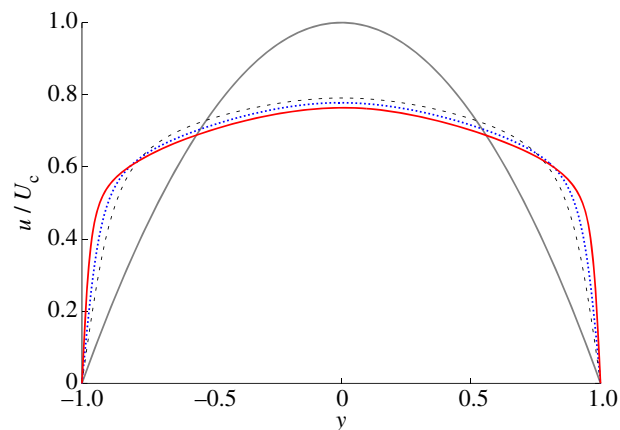


Figure 2. Flow profiles for the dimensionless flow speed as a function of y for the laminar (black solid line) cases (self-similar for all Re) and time-averaged flow profiles for the turbulent cases $(Re, Pe) = (2500, 16750)$, $(4200, 28140)$ and $(10000, 67000)$, as shown by the dashed line, dotted line (blue) and grey (red) solid line, respectively. (Online version in colour.)

equation (2.4) to simulate the translational diffusivity D_t , non-dimensionalized as $D_t^* = D_t/(U_c H)$. These flow-based parameters can be transformed into cell-based non-dimensional parameters for comparison with analytical predictions. From the definitions above and equation (2.13) it can be shown that $Pe = d_r^*/v_s^2$ (flow Péclet number), $\beta = d_r^*/v_s$ (swimming Péclet number) and $\sigma(y) = -(1/3)\chi'(y)/d_r^*$ (local dimensionless shear rate). Since $\chi'(y) = -3y$ for plane Poiseuille flow, the maximum dimensionless shear is given by $|\sigma_{\max}| = 1/d_r^* = \eta\lambda$, where we recall $\lambda = 1/(2Bd_r)$, the non-dimensional bias parameter. Simulations are more readily interpreted and compared with analytical theory in terms of these parameters (shown in table 1).

The dimensionless flow profiles corresponding to the Reynolds numbers used in this study are plotted in figure 2 for the benefit of the reader. Laminar flows are self-similar, so the non-dimensional flow has the same parabolic profile for all Re . Time-averaged turbulent flows have distinct profiles that depend on the Reynolds number. Note that the number of degrees of freedom in the turbulent flow simulations scales as Re^9 [27]. This makes large Re simulations computationally expensive. For this reason, we do not investigate dispersion for flows beyond $Re = 10000$.

3. Results

3.1. Passive tracers: classical dispersion

The dispersion of passive tracers, such as molecular dyes or non-motile cells, is generally well understood. In laminar channel flow passive tracers are transported on average at the mean flow speed; there is no drift relative to mean flow: $A_0 = 0$. The effective axial diffusivity D_e is given at long times by the Taylor–Aris result $D_e = 1 + (8/945)Pe^2$ [3,32]. As a benchmark test, we carried out DNS for passive tracers (solving equation (2.4) with $v_s = 0$ and translational noise to simulate molecular diffusivity; see §2). A typical result is shown in figure 3 for $Pe = 3000$. As expected at long times $A_0 \rightarrow 0$ and the Taylor–Aris diffusivity prediction, $D_e = 76191$, compares very well with simulation results. The skewness slowly tends to zero at long times as $t^{-0.5}$ (see the electronic supplementary material), suggesting approach to a normal distribution [7]. The above results for passive tracers will be compared with the dispersion of gyrotactic swimmers. They can also be used to check our analytical theory in the limiting case of

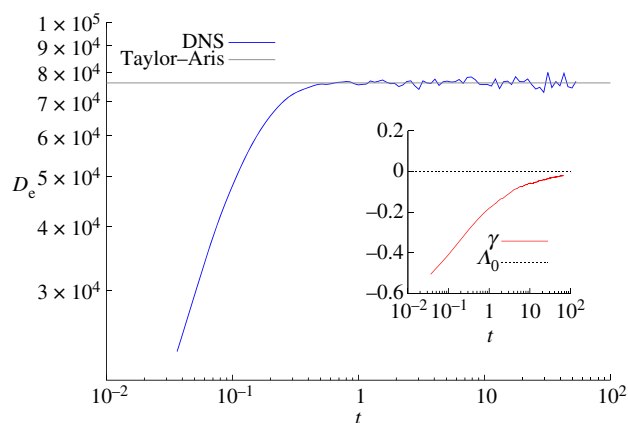


Figure 3. Time dependence of the effective diffusivity, D_e , for passive tracers in a laminar flow with $Pe = 3000$. For long times $D_e \simeq 76191$, the constant value predicted from the classical Taylor–Aris dispersion for passive tracers (see text). The inset shows the expected zero drift, Λ_0 , above the mean flow and the skewness, γ , tending to zero at long times, suggesting an approach to the expected normally distributed axial concentration profile [7]. (Online version in colour.)

unbiased swimmers with no gyrotactic response to flow. In this limit, $\mathbf{q} = 0$ and the diffusivity tensor is isotropic $\mathbf{D} = \mathbf{I}/6$, where \mathbf{I} is the identity matrix (see also [19]). Thus, in the calculation of equations (2.20) and (2.22), $D^{xx} = 1/6 = D^{yy}$, $q_i = 0$ and $D^{xy} = 0$ (as $Y_0^0 = 1$ and $J(y) = 0$), so that $\Lambda_0 = 0$, as expected for passive tracers, and $D_e = 1/6 + 6(8/945)Pe^2$. In the Bees & Croze theory, D_e and Pe are scaled with respect to the diffusivity scale V_s^2/d_r . Using the scaling $(1/3)V_s^2/(2d_r)$ for random diffusers in three dimensions, we recover $D_e' = 1 + (8/945)Pe^2$, the classical Taylor–Aris result for channels.

The dispersion of passive tracers in turbulent channel flows has also been elucidated [8]. Elder derived the (dimensional) effective diffusivity K of passive tracers in turbulent open channel flow as $K = 5.9 u_\tau H$, where u_τ is the friction velocity and H is the channel depth [6,8]. This open channel result applies equally to a closed channel with half-width H . Using the approximation $U_c/u_\tau \approx 5 \log_{10} Re$ [27], this result can be written as $K \approx 2.72vRe/\ln(Re)$. To compare results across the laminar–turbulent transition (see later in figure 12), we use equations (2.7) and (2.13) to obtain a non-dimensional turbulent diffusivity $D_e = K/D_0$ as a function of Pe , where D_0 is the characteristic diffusivity scale. However, we stress that the diffusivity depends only on Re in the turbulent case: molecular diffusivity is negligible [5]. Note also from the result of Elder how the effective turbulent diffusivity grows less sharply with Pe than in the laminar regime. Below, we will contrast the classical predictions for dispersion discussed in this section with our new results for biased swimmers.

3.2. Gyrotactic algae: a new, non-classical ‘swimming dispersion’

3.2.1. Laminar downwelling flows: gyrotactic swimming strongly affects dispersion

It is illuminating to consider how gyrotactic cells distribute across a channel in downwelling laminar flows, as this determines their streamwise dispersion. Figure 4 displays the time evolution of the cross-sectional cell concentration profiles in

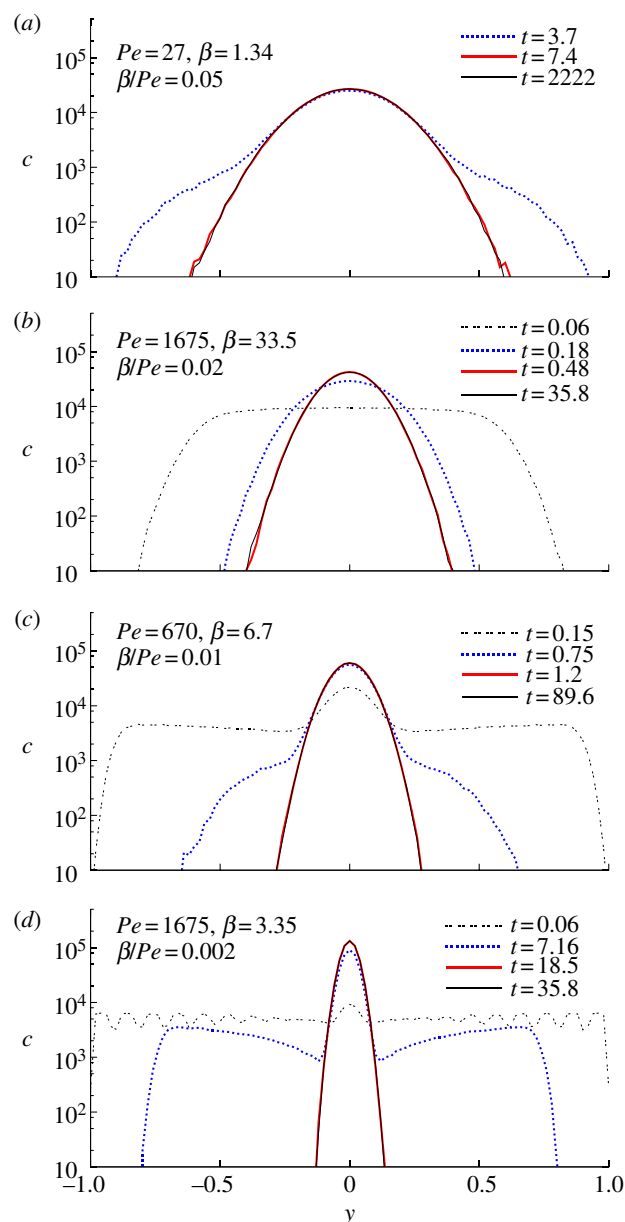


Figure 4. Evolution of the DNS concentration profiles (not normalized) in the wall-normal direction y for gyrotactic cells in downwelling channel flow with $(Pe, \beta) = (a) (27, 1.34)$, $(b) (1675, 33.5)$, $(c) (670, 6.7)$ and $(d) (1675, 3.35)$. At large times, the gyrotactic cells are observed to accumulate around the channel centre. The profiles become more peaked as the ratio β/Pe is decreased (see also figure 5). (Online version in colour.)

the wall-normal direction y for a selection of values of (Pe, β) . It is seen that an initially uniform concentration profile evolves to one focused at the channel centre. This is what one would expect for swimming gyrotactic cells with orientations biased by combined gravitational and viscous torques [11,12]. At the population scale, the long-time concentration distribution is a result of the balance between cross-stream cell diffusion and biased swimming. In §2.3, the cell concentration (normalized by its mean) is given by equation (2.21): $Y_0^0 = c(y)/\bar{c} \propto \exp(\beta \int_0^y (q^y/D_m^{yy}) ds)$. (Recall subscripts $m = G, F$ denote solutions of the GTD and FP microscopic models, respectively.) The two models predict a qualitatively different dependence of concentration distribution on Pe and β . While the FP approach allows for cells to diffuse more and focus less as the cells tumble in flows with large shear rates, the GTD model finds that with increasing local shear rate, σ ,

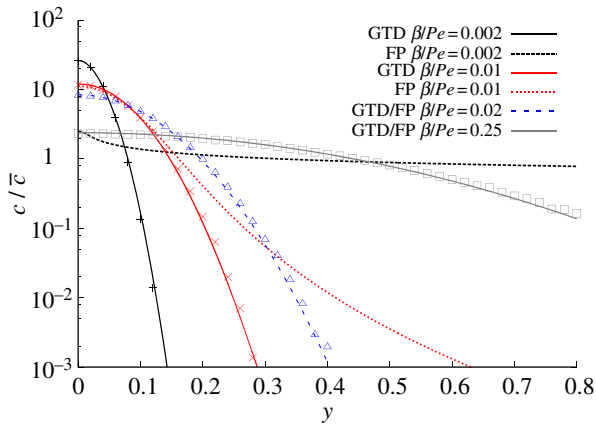


Figure 5. Long-time concentration profiles from DNS, normalized by the mean concentration \bar{c} , with $(Pe, \beta, \beta/Pe)$: (1675, 3.35, 0.002) (plus symbols), (670, 6.7, 0.01) (crosses), (1675, 33.5, 0.02) (triangles), (27, 6.7, 0.25) (squares). Theoretical predictions from the GTD and FP models (lines, see legend) are compared with the simulations. GTD predictions agree very well with DNS profiles for all β/Pe , but for FP agreement is poor for small values of this ratio. As in figure 4, DNS profiles broaden with increasing β/Pe . This is as predicted by GTD, where profiles can be approximated as Gaussians of width $y_0 = [(2/\lambda)(\beta/Pe)]^{0.5}$; see text and appendix A. (Online version in colour.)

the diffusivity of tumbling cells decreases faster than the decrease in cross-stream cell focusing.

It has hitherto been unclear if GTD, mathematically more complicated than the FP approach, provides more accurate predictions. Thus, we compare the DNS results and theoretical predictions from the two models. Figure 5 displays the long-time cell distributions from DNS contrasted with theoretical predictions using GTD and FP. It is clear that DNS profiles get broader with increasing β/Pe , so profiles are shown for different values of this ratio (individual values of Pe and β are shown in the figure caption). GTD and DNS profiles agree very well for all the values of β/Pe simulated. The FP model only agrees well with the DNS for large values of β/Pe , where predictions coincide with those of GTD. The agreement is poor at low values (very poor for the sharply focused distribution $\beta/Pe = 0.002$). It is possible to quantify the scaling of the breadth of the profiles with the ratio β/Pe . From asymptotic expressions for the ratio q^y/D_G^{yy} [19], it follows that GTD profiles can be conveniently approximated by Gaussian distributions of width $y_0 = [(2/\lambda)(\beta/Pe)]^{0.5}$ (see appendix A). Clearly, these predictions need also to be tested experimentally (see §4), but the DNS results indicate that GTD is likely a superior model of gyrotactic response to flow than the FP approach.

As for the passive case of figure 3, we have quantified dispersion using the statistical measures: $\Lambda_0(t)$, the drift above the mean flow; $D_e(t)$, the effective diffusivity; and $\gamma(t)$, the skewness. These are plotted in figure 6 for $Pe = 27$, 670 and 1675 at the fixed value of $\beta = 33.5$. In these simulations, statistically stationary values for $\Lambda_0(t)$ and $D_e(t)$ are achieved for dimensionless times $t \sim 1$; steady values are reached earlier for larger Pe . In terms of the Bees & Croze dispersion theory, steady dispersion is achieved in the long-time limit when transient solutions to the moment equations have died down: $t \gg \tau_1$. The analysis of transient solutions with DNS will be carried out in a future study, but it is reasonable that gyrotaxis makes the approach to steady dispersion faster

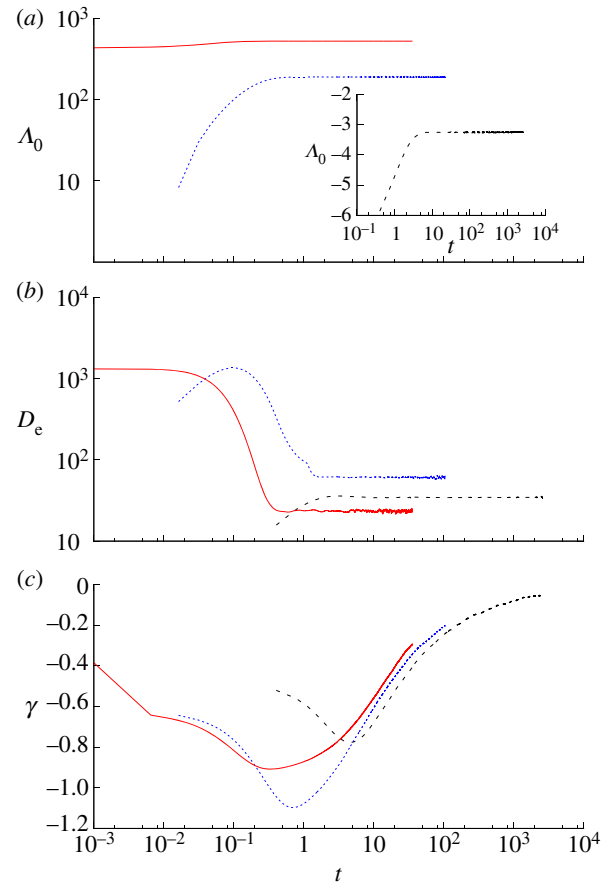


Figure 6. Evolution of the dispersion of a population of gyrotactic swimming algae in a vertical downwelling laminar channel flow. The (a) drift above the mean flow, $\Lambda_0(t)$, (b) effective diffusivity, $D_e(t)$ and (c) skewness, $\gamma(t)$, are displayed for $Pe = 27$ (dashed line), 670 (dotted line) and 1675 (solid line), for fixed $\beta = 33.5$. Uniquely, gyrotactic swimming algae have a non-zero drift above the mean flow, in contrast to the passive tracers in figure 3. Even more peculiarly, $\Lambda_0 < 0$ for $Pe = 27$. This is because of cell upswimming at mid-channel; see text. (Online version in colour.)

than for passive tracers. The skewness is negative and approaches zero (with $\gamma \sim t^{-0.49}$ for $Pe = 27$; see the electronic supplementary material), suggesting a distribution tending to normality. Bees & Croze predicted the power-law decay $\gamma = \gamma_0 t^{-1/2}$ (the pre-factor $\gamma_0(\beta, Pe)$ depends on gyrotactic swimming) with the same exponent as the passive case [7].

The steady gyrotactic swimmer dispersion displays some very surprising features, evident in the data presented in figure 6. For example, Λ_0 , zero in the passive case, grows from a negative value to large positive values as Pe is increased. The effective diffusivity D_e , on the other hand, shows a non-monotonic behaviour for increasing Pe (recall $D_e \sim Pe^2$ for passive tracers). We can qualitatively account for this behaviour considering the concentration distributions analysed above. Cells are biased to swim to the centre of the channel. Here only the torque owing to gravity acts on cells, so cells swim upward at their mean swimming speed, which may be comparable with the mean flow speed for small Pe leading to $\Lambda_0 < 0$. For large Pe , the upward swimming speed is negligible. As cell accumulation at the centre of the channel increases with Pe due to gyrotaxis they drift more relative to the mean flow, and so Λ_0 is an increasing function of Pe . Cell accumulation at the centre of the channel removes

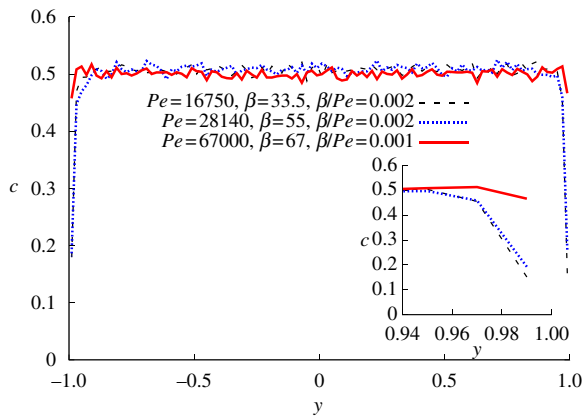


Figure 7. Single realizations of the long-time concentration profiles (not normalized) in direction y for gyrotactic cells in downwelling turbulent flows for $(Pe, Re) = (16\,570, 2500)$, $(28\,140, 4200)$ and $(67\,000, 10\,000)$, as shown. Gyrotaxis causes depletion of cells from regions close to the walls where shear is large, but the mean profile departs little from uniform concentration (expected of passive tracers). (Online version in colour.)

them from regions of high shear rate, eventually leading to a decrease in their effective axial diffusivity with increasing Pe .

The Bees & Croze dispersion theory allows for the quantification of this intriguing dispersion behaviour; the results are summarized in §3.3. Prior to this, we shall consider time-dependent dispersion in turbulent flows.

3.2.2. Turbulent downwelling flows: persistent but weaker swimming dispersion

Here, we describe the first DNS study of the dispersion of gyrotactic cells in turbulent channel flows. To compare turbulent and laminar results we first focus on downwelling flows. Simulations were performed for $(Pe, Re) = (16\,570, 2500)$, $(28\,140, 4200)$ and $(67\,000, 10\,000)$, with corresponding values of $\beta = 33.5, 55$ and 67 . The long-time wall-normal cell concentration profiles for these turbulent flows are shown in figure 7. It is clear that the mean cell concentration is uniform (barring small fluctuations about the mean) for almost the entire channel width, except for regions close to the wall that are gyrotactically depleted of cells (gyrotactically depleted regions occupy only a small fraction (less than 4%) of the channel width). Shear and advection experienced by a cell in turbulent flows are very different from the laminar case. The turbulent flow can be thought of as a time-averaged base profile on which are superposed turbulent fluctuations (the well-known Reynolds decomposition [27]). The shear rate of the base profile is close to zero in the middle of the channel, and large at the walls (figure 2) and deviates from the laminar case; this alone will lead to broader concentration profiles. On top of this, turbulent fluctuations perturb the flow, causing cell reorientation and advection. We can think of turbulence as endowing cells with an increased diffusivity [35] acting to make gyrotactic accumulations in this downwelling case less pronounced. Only close to the walls is the impact of mean shear sufficient to cause significant cell depletion; an effect that increases with β/Pe in a similar fashion to the laminar case. The small but measurable effects of gyrotaxis on the concentration profiles are reflected in the time-dependent dispersion measures for the turbulent case, plotted in figure 8 for the same values of (Pe, β) considered above. We leave a detailed analysis of

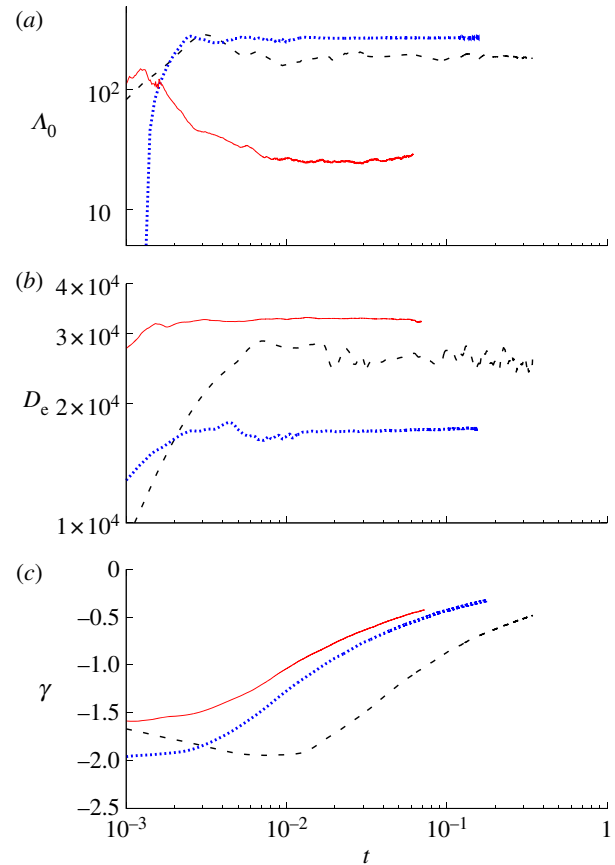


Figure 8. Evolution of statistical measures ((a) to (c)), as for the laminar case for gyrotactic cells in a vertical downwelling turbulent channel flow for $(Pe, Re) = (16\,570, 2500)$, $(28\,140, 4200)$ and $(67\,000, 10\,000)$, as shown by the dashed line, dotted line and solid line, respectively. The long-time values of the drift above the mean flow, Λ_0 , and effective diffusivity, D_e , are plotted in figure 12. Values of β are not shown for clarity (see table 1). (Online version in colour.)

transients to a future study but note that, because of the increased diffusivity by turbulence, the approach to the limiting behaviour is faster than in the laminar case. Less obviously, the long-time dispersion retains a rich behaviour (observe the order of the curves in figure 8). In particular, note that cells have a non-zero drift Λ_0 ; this is because of local focusing of cells in downwelling regions of the fluctuating flow. The dispersion of gyrotactic swimmers is thus qualitatively distinct from that of passive tracers even in a turbulent flow. As Pe is increased to the maximum value simulated, Λ_0 decreases while D_e increases, indicative of increased mixing by turbulence.

3.2.3. Dispersion in upwelling flows

The case of dispersion in flows directed vertically upwards (against the direction of gravity) is considered here briefly. The DNS results for the same values of Pe and β as the laminar downwelling case of §3.2.1 are shown in figure 9. The drift above the mean flow Λ_0 is positive for small Pe , and grows more negative with increasing Pe . This behaviour is the result of the peculiar distribution of gyrotactic cells in the flow: cells in upwelling flow are biased to swim not to the channel centre, but to the walls [11]. Interestingly, accumulation and dispersion depend critically on the flow direction! The inset of figure 9b shows the cell concentration c in the wall-normal direction y , demonstrating the wall

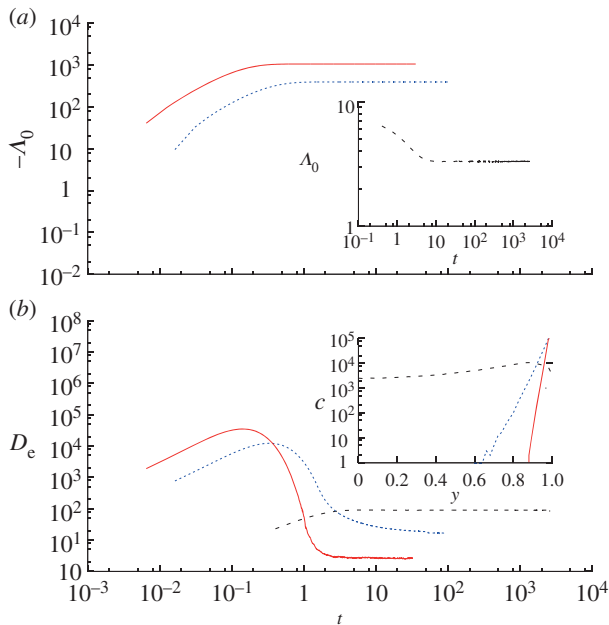


Figure 9. Dispersion of gyrotactic algae in vertical upwelling laminar flows. The parameters are as for the downwelling laminar case ($\beta = 33.5$ and $Pe = 27, 670$ and 1675 denoted by the dashed line, dotted line and solid line, respectively). (a) The drift $-\Lambda_0$ (inset: the positive drift for $Pe = 27$). (b) The effective diffusivity D_e (inset: the concentration profiles). (Online version in colour.)

accumulation, which becomes more peaked with decreasing β/Pe (for more details, see the electronic supplementary material). Thus, for fixed β and increasing Pe , cells focus at the walls and experience slower flow and less of the shear profile, leading to a decrease in both the drift, Λ_0 , and the diffusivity, D_e . The change of sign in Λ_0 has a similar explanation as for the downwelling case.

The upwelling turbulent case, presented in figure 10, was also investigated for the same parameter values as the downwelling case of §3.2.2. We see that dispersion measures do not reach steady values: both Λ_0 and D_e grow monotonically with time for the duration of the simulation run. The inset in figure 10a shows the cell concentration profiles displaying a thin band of gyrotactic accumulation at the walls. Cells that end up in this band are subject to strong dispersal by the large mean shear rate at the wall. Diffusion, whether owing to turbulence or swimming, is not strong enough to balance the shear-induced migration towards the wall, as is the case for laminar flow, leading to non-steady dispersion over the course of the simulations.

However, the upwelling dispersion results presented above are not realistic for swimming species that are negatively buoyant; accumulations of negatively buoyant cells at the walls can result in instability and the formation of downwelling flow and descending plumes near the walls [11]. These instabilities will differ in their extent in the laminar and turbulent upwelling cases, but we expect buoyancy-driven wall flows to ensue after accumulation in both regimes.

3.2.4. The effects of cell elongation

So far we have assumed that the gyrotactic cells are spherical, setting the eccentricity parameter α_0 to zero. Here, we present simulations obtained with a non-zero eccentricity, $\alpha_0 = 0.8$, for the cases $(Pe, \beta) = (670, 6.7)$ (laminar) and $(28\,140, 55)$ (turbulent). We chose the relatively large value of $\alpha_0 = 0.8$ to

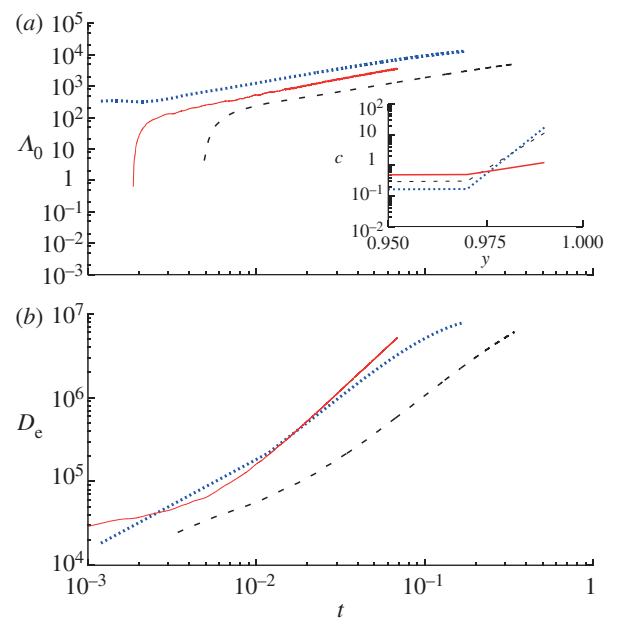


Figure 10. Dispersion in upwelling turbulent flows for the same parameters as the downwelling case. (a) Mean drift $-\Lambda_0$ (inset: cell concentration profile close to the wall). (b) Effective diffusivity D_e . Note that in the turbulent case neither of the statistical measures reaches a steady value. $(Pe, Re) = (16\,570, 2500), (28\,140, 4200)$ and $(67\,000, 10\,000)$, as shown by the dashed line, dotted line and solid line, respectively. (Online version in colour.)

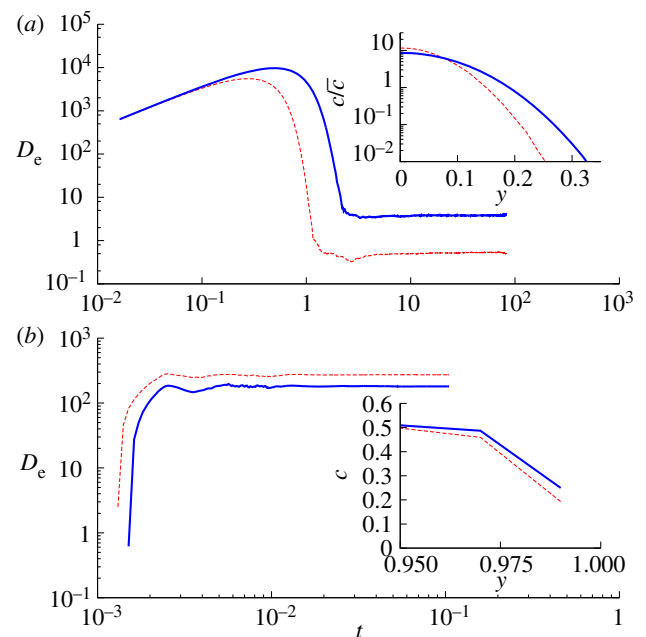


Figure 11. The effect of cell elongation on dispersion. Simulations of the effective diffusivity D_e performed for spherical ($\alpha_0 = 0$, dashed line) and elongated ($\alpha_0 = 0.8$, solid line) cells for downwelling flows. (a) Laminar flow, with $Pe = 670$ and $\beta = 6.7$; (b) turbulent flow, with $Pe = 28\,140$ and $\beta = 55$. Even for the large α_0 value considered, qualitative effects are not observed and significant quantitative differences are apparent only in the laminar case (see text and electronic supplementary material). (Online version in colour.)

emphasize the effect of eccentricity. Results for the effective diffusivity for downwelling flow are shown in figure 11 for the laminar and turbulent cases. The insets in the figure display the concentration profiles. We see that in the laminar case the effect of eccentricity is to broaden the profile, as

observed in Bearon *et al.* [36]. This broader distribution results in an increased value of effective diffusivity (cells sample more of the shear profile). In the turbulent case of figure 11*b*, there is a much smaller broadening effect. The effect of cell elongation on other statistical measures for the parameters considered here is marginal (see the electronic supplementary material). If realistic values of biflagellate eccentricity ($\alpha_0 = 0\text{--}0.3$ [13]) are used, predictions for the dispersion of biased swimmers are not very different from those obtained here for spherical cells.

3.3. Long-time dispersion of gyrotactic cells as a function of Pe (Re) across the laminar–turbulent transition

Having analysed the full time dependence of gyrotactic cell dispersion, we concentrate here on its long-time behaviour. In this limit, laminar DNS results can be compared with predictions from analytical theory for drift Λ_0 and diffusivity D_e as functions of Pe (for given β). The theory requires as inputs expressions for the mean swimming direction, \mathbf{q} , and diffusivity tensor, \mathbf{D} , obtained from microscopic stochastic models. We test two alternative microscopic models: FP and GTD. The models predict qualitatively different functional forms for the components of \mathbf{D} as a function of the dimensionless shear σ . Correspondingly, predictions for $\Lambda_0(Pe)$ and $D_e(Pe)$ obtained with the two models also differ.

The predictions from the analytical theory are shown as curves in figure 12*a,b*, for fixed values of β ; the corresponding DNS results for selected Pe and the same values of β are shown as points. It is clear that, for the GTD predictions, the agreement between theory and simulation for Λ_0 is very good. The GTD prediction is that, for fixed β , Λ_0 increases with Pe , tending to a linear behaviour for large enough Pe . This is in contrast to passive tracers, for which $\Lambda_0 = 0$. For small Pe the FP prediction coincides with that of GTD, but, for larger Pe , Λ_0 tends to a constant instead of growing with Pe . The smaller the value of β , the greater is the difference between the predictions of the two microscopic models (as for concentration profiles). A similar trend is seen for the effective diffusivity D_e , shown as a function of Pe in figure 12*b*. We see that for $\beta = 33.5$, both GTD and FP predict a diffusivity rising and then falling with Pe , consistent with DNS results. For smaller values of β , however, the FP and GTD predictions are different: GTD predicts a decrease of D_e at large Pe , while FP predicts a (second) rise. The DNS results are not compatible with this rise, but agree well with the GTD prediction. We remark that this dispersion behaviour is unique to swimming cells. It is due to gyrotactic bias and the ensuing accumulations that change the distribution of cells and thus their dispersion, as discussed above. Also shown in figure 12*b* is the diffusivity for passive tracers, which grows as $D_e \sim Pe^2$ [4], without the decrease at large Pe we predict for gyrotactic swimmers.

In earlier sections, we have seen how the effect of gyrotaxis on accumulation and dispersion is more subtle in turbulent flows. DNS results for long-time dispersion reflect these changes and are also summarized in figure 12. An analytical theory of turbulent swimming dispersion has yet to be formulated, so we compare only with theory for passive tracers. It is seen for all simulations with $\beta = 33.5$ that the value of Λ_0 , which was growing with Pe in the laminar

regime, drops a little just beyond the transition to turbulence. On the other hand, the diffusivity D_e , which was falling at large laminar Pe , suddenly acquires a sizeable value. The turbulent shear profile and fluctuations alter the distribution of cells. The effect of gyrotaxis is thus weakened but still measurable in the dispersion, which is similar but not identical to that of passive tracers. The fractional drift above the mean flow speed $U = (2/3)U_c$ is given by $\delta \equiv \lim_{t \rightarrow \infty} (1/U)[dm_1/dt - U] = (3/2)\Lambda_0/Pe$; this is rather small for large turbulent Pe values. Nevertheless, it is remarkable that drift should not be zero in a turbulent flow, as it is for passive tracers. For very large values of Pe , we expect gyrotaxis to have practically no effect on dispersion: the time-averaged concentration profile will be well approximated as uniform and $\Lambda_0 = 0$. Indeed, our results suggest that $\Lambda_0 \rightarrow 0$ for increasing Pe in the turbulent regime.

4. Discussion

We have studied the dispersion of gyrotactic swimming algae in channel flows by DNS and analytical theory. This is the first study to evaluate cell distributions and statistical measures of gyrotactic cell dispersion (drift above mean flow Λ_0 , effective diffusivity D_e and skewness γ) for flows on either side of the laminar–turbulence transition. We find unique cell accumulations and dispersion with non-zero drift and a non-monotonic diffusivity with Pe , with qualitative differences for upwelling and downwelling flows. The dispersion behaviour is remarkable and unique to gyrotactic swimming cells: passive tracers are transported at the mean flow rate ($\Lambda_0 = 0$), the diffusivity increases with Pe ($D_e \sim Pe^2$) [4], and dispersion is insensitive to channel orientation. In the laminar downwelling regime, simulation results were compared with the predictions derived from a recently formulated general analytical theory of swimming dispersion [18,19], applied here for the first time to channel geometry. The theory requires as inputs expressions for cell response to the local shear in a flow, determined from microscopic models.

We have evaluated predictions based on two alternative models: the FP [33,34] and GTD [19–21] approaches. Which model is more realistic has long been a matter of debate [20]. Here, we find that the DNSs are in excellent agreement with analytical predictions based on GTD, providing the first evidence that GTD is much better than the FP model at describing swimmers in flows. As well as validating analytical predictions for laminar flow, the DNSs allow us to explore the industrially relevant dispersion of gyrotactic algae in turbulent channel flows. In the turbulent regime, the effects of gyrotaxis (accumulations resulting in non-zero drift) persist, but are much more subtle. Effective diffusivity in downwelling turbulent flows is similar to that of passive tracers. These are the first full DNSs of biased swimmers in turbulent channel flows; previous studies having concentrated on vortical flow [37] or synthetic approximations of homogeneous isotropic turbulence [31,35].

The fact that swimming algae in channel flows distribute very differently to passive tracers has important practical consequences for the culture of swimming species. The most dramatic implications of our findings are for photobioreactors that operate using laminar flows. For example, in draft tube air-lift bioreactors, bubbles up a central draft

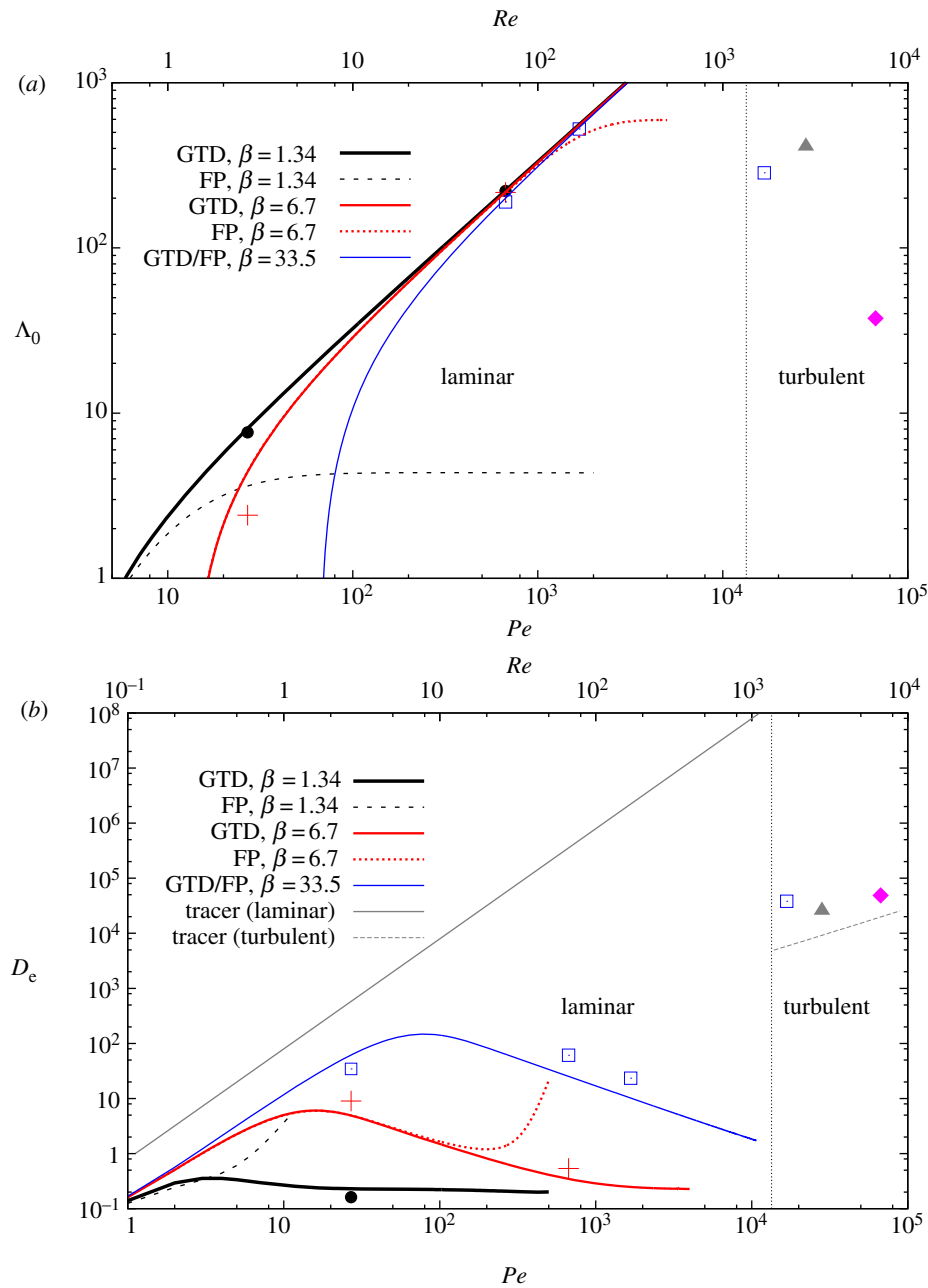


Figure 12. DNS results for long time (a) drift, Δ_0 , and (b) diffusivity, D_e , as a function of Pe for gyrotactic cells in downwelling flows. Values of β are 1.34 (circles), 6.7 (plus symbols), 33.5 (squares), 55 (triangles), 67 (diamonds). The DNS results are compared with predictions from the Bees & Croze dispersion theory, using the FP and GTD microscopic models, plotted as lines for the same β values as the DNS; see legend. Simulation results compare well with GTD predictions for all β , but, for $Pe \gtrsim 1$, they are incompatible with FP predictions for small β . Also plotted in (b) are the classical results for passive tracers. In laminar flows, $\Delta_0 = 0$ and $D_e \sim Pe^2$, in sharp contrast with our predictions for swimmers. In turbulent flow, we still have $\Delta_0 > 0$ for swimmers, though it appears to decay to zero with increasing Pe . The turbulent D_e for swimmers is close to the passive case; see text. (Online version in colour.)

tube (riser) mix and aerate cells, which then circulate down the channel formed between the draft tube and the surface of the reactor (downcomer; figure 1c). The Bees & Croze [18] analytical theory, confirmed by simulations, predicts that gyrotactic swimming algae will be focused more and more sharply at the centre of the downcomer as Pe (the flow rate for fixed channel width, or Re) is increased. For example, considering a flow with $Pe = 1675$ ($Re = 250$) and the swimming Péclet number $\beta = 33.5$ ($H = 5$ cm), we predict that the concentration at the walls is a vanishingly small fraction of the mean, given by $c/\bar{c} \approx \exp[-Pe\lambda/(2\beta)]/\sqrt{\pi\beta/(2\lambda Pe)} \sim 10^{-23}$ (using $\lambda = 2.2$ for *C. augustae*; see approximations in appendix A). Non-swimming cells and molecular solutes, on the other hand, would be uniformly distributed across the tube, $c/\bar{c} = 1$. As we can reasonably assume

that the probability of cell adhesion to the walls is proportional to the concentration there, we predict that surface fouling by gyrotactic cells will be markedly reduced relative to non-swimming cells. Fouling can be a big problem in closed bioreactors because cell build-up can prevent light penetration and thus growth, and in extreme cases can clog reactor conduits [38].

The peculiar dispersion of biased swimmers will also affect growth in bioreactors. Our results indicate that gyrotactic swimmers in a downwelling flow drift faster and diffuse less than passive tracers, which travel at the mean flow. We can make experimentally measurable predictions for the dispersion of *C. augustae*, the alga whose swimming parameters (reorientation time B , rotational diffusivity d_r , swimming speed V_s) we have used in this study. For example, in the case with $\beta = 6.7$ and $Pe = 670$ (e.g. a realistic, small air-lift

Table 2. Parameters used in the functional fits of solutions to the FP and GTD models used for our analytical theory predictions (from [19]).

	a_0	a_2	a_4	b_2	b_4
\mathbf{a}^x	5.70×10^{-1}	3.66×10^{-2}	0	1.75×10^{-1}	1.25×10^{-2}
\mathbf{a}^y	2.05×10^{-1}	1.86×10^{-2}	0	1.74×10^{-1}	1.27×10^{-2}
\mathbf{a}_G^{xx}	5.00×10^{-2}	1.11×10^{-1}	3.71×10^{-5}	1.01×10^{-1}	1.86×10^{-2}
\mathbf{a}_G^{yy}	9.30×10^{-2}	1.11×10^{-4}	0	1.19×10^{-1}	1.63×10^{-4}
\mathbf{a}_G^{xy}	9.17×10^{-2}	1.56×10^{-4}	0	2.81×10^{-1}	2.62×10^{-2}
\mathbf{a}_F^{xx}	5.60×10^{-2}	3.23×10^{-2}	1.70×10^{-5}	2.70×10^{-1}	1.42×10^{-4}
\mathbf{a}_F^{yy}	9.30×10^{-2}	5.73×10^{-4}	1.85×10^{-3}	4.96×10^{-2}	1.54×10^{-2}
\mathbf{a}_F^{xy}	1.58×10^{-2}	0	0	9.61×10^{-2}	7.88×10^{-2}

with $H = 1$ cm and $U_c = 1$ cm s $^{-1}$) the fractional drift above the mean flow, $\delta = (3/2)\Lambda_0/Pe$, is estimated to be $\delta = 0.48$. In other words, *C. augustae* cells drift about 50 per cent faster than passive tracers, such as nutrients. For the non-dimensional effective diffusivity we predict $D_e = 0.8$: the axial diffusivity of cells is smaller than the cell diffusivity scale ($V_s^2/d_r = 1.49 \times 10^{-3}$ cm 2 s $^{-1}$) in the absence of flow. To compare these predictions with those for passive molecular solutes, we consider the dispersion of CO $_2$ (molecular diffusivity $D_t = 1.6 \times 10^{-5}$ cm 2 s $^{-1}$), a vital nutrient for algae. In the flow under consideration, cells have $Pe = 670$, but CO $_2$, with its smaller diffusivity, has $Pe_t = PeV_s^2/(d_r D_t) = 6.2 \times 10^4$. Thus, the non-dimensional tracer diffusivity is $D_{e,t} \approx (8/945)Pe_t^2 \sim 10^7$, from the Taylor–Aris result. Re-dimensionalizing swimming and solute effective diffusivities by multiplying by diffusivity scales, we find that the axial diffusivity of CO $_2$ along a channel is approximately 10^5 times greater than that of *C. augustae*. This dramatic differential dispersion of cells and nutrients could have important consequences for swimming cell growth in reactors.

In turbulent downwelling flows, which may also be realized in air-lifts, the effects of gyrotactic swimming are much less pronounced. Gyrotactic depletion is not as efficient as in laminar flows: the concentration at the walls is at best half of the mean concentration. We thus expect significant fouling in air-lifts under turbulent regimes. The influence of gyrotaxis on dispersion is also weaker than in the laminar case. For *C. augustae* in a turbulent channel flow with $\beta = 55$ ($H = 8.4$ cm) and $Pe = 28\,140$ ($Re = 4200$, $U_c = 5$ cm s $^{-1}$), the DNSs predict a fractional drift of $\delta = 0.015$. This is very small, and may be neglected for short channels. As shown in figure 12*b*, the effective diffusivity is of the same order as that of a passive tracer such as CO $_2$.

The excellent agreement between DNS and predictions for dispersion obtained using GTD theory provides a first confirmation that the swimming dispersion theory of Bees & Croze [18] is robust. If they are to be useful in bioreactor engineering design, however, the theoretical and numerical predictions for dispersion need to be tested against experiments. Work is in progress to test GTD predictions for pipe flow with the alga *Dunaliella* [23]. It would also be interesting to test experimentally the numerical and analytical predictions for channel flow presented herein, exploring in particular the laminar–turbulent transition region. Since tubes are often arranged horizontally in bioreactor designs, it will be interesting to study the effect of tube orientation on biased swimmer

dispersion. Croze *et al.* [39] have carried out experiments on the dispersion of *C. augustae* in horizontal pipe flow for low Pe . They found a complex transport mediated by sheared bio-convection patterns and suggested that such cell-driven flows could alter the transition to turbulence. It would be interesting to study this transition experimentally and with the aid of simulations such as presented here, including the effects of cell buoyancy (especially for small Pe flows) [19].

More generally, our study could be extended to open channel flows, such as those present in pond bioreactors, channels, waterfalls and rivers. The swimming dispersion effects we have explored must also exist whenever biased swimmers are subject to shear in the ocean and lakes. Significantly, about 90 per cent of species implicated in the formation of harmful algal blooms swim using flagella [40]. A recent study has shown that *Heterosigma akashiwo* is gyrotactic and could be responsible for thin layer formation [15]. However, the role of biased swimming in the dispersion and ecology of algae remains largely unexplored.

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Appendix A. Microscopic model solutions

A.1. Numerical solutions for the Fokker–Planck and generalized Taylor dispersion models

$\mathbf{q}(\sigma)$ and $\mathbf{D}_m(\sigma)$ predicted by the FP and GTD microscopic models have the following structure:

$$\mathbf{q} = \begin{bmatrix} q^x(\sigma) \\ q^y(\sigma) \\ 0 \end{bmatrix} \quad \text{and} \quad \mathbf{D}_m = \begin{bmatrix} D_m^{xx}(\sigma) & D_m^{xy}(\sigma) & 0 \\ D_m^{xy}(\sigma) & D_m^{yy}(\sigma) & 0 \\ 0 & 0 & D_m^{xx}(\sigma) \end{bmatrix}, \quad (\text{A } 1)$$

where we recall that $m = F$ represents solutions of the FP model and $m = G$ those of the GTD model. Such solutions can be obtained using approximations via a Galerkin method [19]. It is convenient to fit such solutions with the

following expressions: $q^x(\sigma) = -P(\sigma; \mathbf{a}^x, \mathbf{b}^x)$; $q^y(\sigma) = -\sigma P(\sigma; \mathbf{a}^y, \mathbf{b}^y)$; $D_m^{xx}(\sigma) = P(\sigma; \mathbf{a}^{xx}, \mathbf{b}^{xx})$; $D_m^{yy}(\sigma) = P(\sigma; \mathbf{a}^{yy}, \mathbf{b}^{yy})$; $D_m^{xy}(\sigma) = -\sigma P(\sigma; \mathbf{a}^{xy}, \mathbf{b}^{xy})$. Here, the rational function $P(\sigma; \mathbf{a}, \mathbf{b})$ is given by

$$P(\sigma; \mathbf{a}, \mathbf{b}) = \frac{a_0 + a_2 \sigma^2 + a_4 \sigma^4}{1 + b_2 \sigma^2 + b_4 \sigma^4}, \quad (\text{A } 2)$$

and, for $\lambda = 1/(2d_r B) = 2.2$, the coefficients \mathbf{a} and \mathbf{b} are provided in table 2.

A.2. Approximate generalized Taylor dispersion profiles and width scaling

Using asymptotic results derived in [19], we derive an approximation for the concentration profiles predicted by the GTD model. These profiles are given by equation (2.21) as $c(y)/\bar{c} = \exp[\beta \int_0^y (q^y/D_m^{yy}) ds] / \int_{-1}^1 (1/2) \exp[\beta \int_0^y (q^y/D_m^{yy}) ds] ds'$, where $m = G$ for GTD predictions. For $\sigma \ll 1$ at leading order the GTD prediction asymptotes to

$q^y(\sigma) = -\sigma J_1/\lambda$, where J_1 is a known constant for $\lambda = 2.2$. In the same limit, $D_G^{yy}(\sigma) = J_1/\lambda^2$, so that $\sigma \gg 1$. For $\sigma \gg 1$ at leading order, $q^y(\sigma) = -(2/3)\lambda/\sigma$ and $D_m^{yy}(\sigma) = d_1/\sigma^2$, where $d_1 = 0.68$ for $\lambda = 2.2$. Thus, $q^y/D_G^{yy} \approx -\sigma\lambda$ is a reasonable approximation for all σ . Recalling for channel flow $\sigma = (Pe/\beta^2)y$, we see that $\beta q^y/D_G^{yy} \approx -\lambda(Pe/\beta)y$. Inserting this expression in the equation above and integrating gives the Gaussian profile

$$c(y) \approx \frac{\exp[-(y/y_0)^2]}{\sqrt{\pi/4y_0 \operatorname{erf}(1/y_0)^{y_0 \ll 1}}} \approx \frac{\exp[-(y/y_0)^2]}{\sqrt{\pi/4y_0}} \quad (\text{A } 3)$$

where

$$y_0 \approx \left[\left(\frac{2}{\lambda} \right) \frac{\beta}{Pe} \right]^{0.5} \quad (\text{A } 4)$$

is the width of the profile. This scaling is observed in the concentration profiles obtained from DNS; see the main text.

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